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## Background Information

Petunia possesses Solanaceae-type self-incompatibility (SI), which allows pistils to reject self-pollen preventing inbreeding, but accept non-self pollen for outcrossing. Self/non-self recognition is regulated by the polymorphic $S$-locus (Fig. 1). A single gene at the $S$-locus, $S$-RNase, encodes the pistil specificity determinant. The first S-locus-F-box (SLF) gene, named SLF1, was identified by sequencing a $328-\mathrm{kb}$ BAC contig containing $\mathrm{S}_{2}-$ RNase. Pollen transcriptome analysis revealed 16 additional SLF genes linked to the $S_{2}$-locus (Fig. 1). All 17 SLF genes collectively encode the pollen-specificity determinant.
$S_{2}$-locus
Figure 1. Schematic diagram of $S_{2}$-Iocus. Except for the location of $S_{2}$-SLFF1, the locations of the other $16 S L F$ genes ( $S_{2}$-SLF2 to $S_{2}$-SLFF1) relative to $S_{2}$-RNase are yet undetermined.


Figure 2. Using gene markers to screen the $S_{2} S_{2}$ BAC library. (A). Schematics showing the "pooling" strategy used for screening the $S_{2} S_{2}$ BAC library. (B) An example showing the use of the $\mathrm{S}_{2}$-SLFE Specific primers to screen the library, and the identification of a BAC clone 143020 , containing $\mathrm{s}_{2}$-LLF8.

## Methods

We used SLF2 to SLF17 as markers to isolate BAC clones from the previously constructed $S_{2}$ library (Fig. 2), and used Illumina Miseq and PacBio SMRT sequencing technology to sequence genomic DNA inserts of these BAC clones, as well as of a previously assembled $881-\mathrm{kb}$ BAC contig containing the $328-\mathrm{kb}$ region. The sequence of each BAC clone was assembled using a combination of Illumina MiSeq and PacBio sequence reads. Illumina read processing and assembly was performed in-house, whereas all PacBlo read quality processing was performed through the SMRT analysis pipeline (v2.3.0) (Fig. 3A).


Figure 3: (A). The $S_{2}$-locus assembly workflow. (B) Scaffold statistics of successive assembly steps of $S_{2}$-SLF12. The top panel shows the increase in assembly quality with each successive step, as indicated by comparing the N50 value to the total number of base pairs
assembled (left to right).

## Results

A total of 3.1 Mbp (a single contig each for 13 of the 17 SLF genes) were assembled (Table 1), $20.34 \%$ of which were repetitive sequences (Fig. 4).


No additional SLF genes were discovered, but 38 additional genes were predicted: 30 of unknown origin and 8 annotated as encoding proteins functioning in pollen germination, pollen tube growth or guidance (Table 2). The sequence of the $S_{6 a}$-locus of $P$. inflata, extracted from the draft genome sequence, contained 29 of these 38 genes, indicating shared characteristics between different $S$-loci of the same species.

| Uniprot Accession Number |  |  |  |
| :---: | :---: | :---: | :---: |
| $\begin{aligned} & \hline \text { CNGC } \\ & \text { CNGC9_ARATH } \end{aligned}$ | cyclic nucleotide-gated ion channel | pollen tube growth | Wang et al., (2013) |
| BECN1 | Beclin 1 like protein | ollen germinatio |  |
| $\begin{aligned} & \text { DRM } \\ & \text { DRMI_ARATH } \\ & \hline \end{aligned}$ | DNA (cytosine-5)methyltransferase DRM2 | DNA methylation | Calarco et al., (2012 |
| $\underset{\text { PDI DATGL }}{\text { PDI }}$ | protein disulfide isomerase | pollen tube guid | Wang et al., (200 |
| $\begin{aligned} & \hline \text { R27A } \\ & \text { R27A2_ARATH } \\ & \hline \end{aligned}$ | 60 S ribosomal protein L27a- <br> 2 | protein synthe | Klinge et al., (2011) |
| $\begin{aligned} & \hline \text { WSD } \\ & \text { WSDI_ARATH } \end{aligned}$ | O-acyltransferase | cuticular wax biosynthesis | Let al., (2008) |
| DUL4 | CULLIN-4 | ubiquitination SCF complex member | Seo et al., (2014) |
| $\begin{aligned} & \text { TSS } \\ & \text { TSS_ARATH } \end{aligned}$ | TPR-domain suppressor of STIMPY | cell cycle regulator | ylar et al., (201 |
| Table 2. Eight genes identified in the $S_{2}$-locus of Petunia inflata and annotated as encoding proteins functioning in pollen germination, pollen tube growth, or guidance. . |  |  |  |
| S-locus remnants on Chromosome 1 of self-compatible tomato (Solanum lycopersicum) and potato (Solanum tuberosum) cultivars contained the S-RNase remnant and SLF remnants in a sub-centromeric region, but did not contain any of the 38 annotated genes, suggesting the unique feature of the S-locus genes involved in SI. |  |  |  |



Figure 5. Comparative analyses of the Petunia $S_{2}$-locus and chromosome 1 of both Potato and Tomato. Panel A shows the locations of SLF remnants, as determined by a hiddenmarkov model, in chromosome 1 of Tomato (left), and the locations of the homologs of the 38 genes identified in the Petunia $S_{2}$-locus. Panel B shows a similar analysis using chromosome 1 of Potato. In both panels, the hit density (genes per 500 Kbp ) and locations


Figure 6. Phylogenetic relationships of SLF genes of $S_{2}$-locus and $S_{6 a}$-locus of Petunia inflata and of Petunia axillaris.

For both $S_{2}$ and $S_{6 a}$ loci, comparison of the upstream and downstream non-coding sequences of different SLF genes (Fig. 6 and Fig. 7) revealed that both recombination and retrotransposition might have played a role in the expansion of SLF genes.


Figure 7. Comparison of non-coding sequences flanking SLF genes. Sequence comparison of 10 kbp upstream (top) and 10 Kbp downstream (bottom) between $S_{2}$-SLF12 and $S_{2}$-SLF16 (black line), and between $S_{2}$-RNase and all 17 SLF genes (red line).

## Conclusion

The sequence of the $S_{6 a}$-locus of $P$. inflata, extracted from the draft genome sequence, contained 29 of the 38 genes, indicating shared characteristics between different $S$-loci of the same species. $S$-locus remnants on chromosome 1 of self-compatible tomato (Solanum lycopersicum) and potato (Solanum tuberosum) cultivars contained the S-RNase remnant and SLF remnants in a sub-centromeric region, but did not contain any of the 38 annotated genes, suggesting the unique feature of the $S$-locus genes involved in SI For both $S_{2}$ and $S_{6 a}$ loci, comparison of the upstream and downstream non-coding sequences of different SLF genes revealed that both recombination and retrotransposition might have played a role in the expansion of SLF genes

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## References

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