

Lihua Wu¹, Justin S. Williams², Ning Wang¹, Wasi A. Khatri², Daniele San Román², Teh-hui Kao^{1,2,*}

¹Intercollege Graduate Degree Program in Plant Biology, The Pennsylvania State University, University Park, Pennsylvania 16802 USA

²Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, Pennsylvania 16802 USA

Background Information

Self/non-self recognition is regulated by the polymorphic *S*-locus; matching of the pollen *S*-haplotype with one of the two pistil *S*-haplotypes results in inhibition of pollen tube growth. The *S*-locus houses *S*-RNase for pistil specificity, and, for both *S*₂- and *S*₃-haplotypes, 17 *S*-locus *F*-box (*SLF*) genes for pollen specificity. All *SLF*s are assembled into similar SCF complexes, containing Rbx1, pollen-specific Cullin1 and Skp1-like protein. According to the collaborative non-self recognition model, for a given *S*-haplotype, each SCF complex interacts with a subset of non-self *S*-RNases to mediate their ubiquitination and degradation by the 26S proteasome. Our lab has used a transgenic assay (Fig. 1) to determine interaction relationships of *SLF* proteins and *S*-RNases. Among those determined, *S*₂-*SLF*1 and *S*₃-*SLF*1 (an allelic pair of *SLF*1, differing in 44 amino acids) show differential interactions with several *S*-RNases, i.e., *S*₂-*SLF*1, but not *S*₃-*SLF*1, interacts with *S*₃⁻, *S*₇⁻ and *S*₁₃⁻-RNases (Table 1).

	<i>S</i> ₂ ⁻ RNase	<i>S</i> ₃ ⁻ RNase	<i>S</i> ₅ ⁻ RNase	<i>S</i> _{6a} ⁻ RNase	<i>S</i> ₇ ⁻ RNase	<i>S</i> ₁₁ ⁻ RNase	<i>S</i> ₁₂ ⁻ RNase	<i>S</i> ₁₃ ⁻ RNase	<i>S</i> ₁₆ ⁻ RNase	<i>S</i> ₂₁ ⁻ RNase	<i>S</i> ₂₄ ⁻ RNase
<i>S</i> ₂ - <i>SLF</i> 1	—	+	—	—	+	—	+	+	—	—	—
<i>S</i> ₃ - <i>SLF</i> 1	—	—	—	—	—	—	+	—	—	—	—

Table 1. *S*₂-*SLF*1 and *S*₃-*SLF*1 show differential interactions with three *S*-RNases.

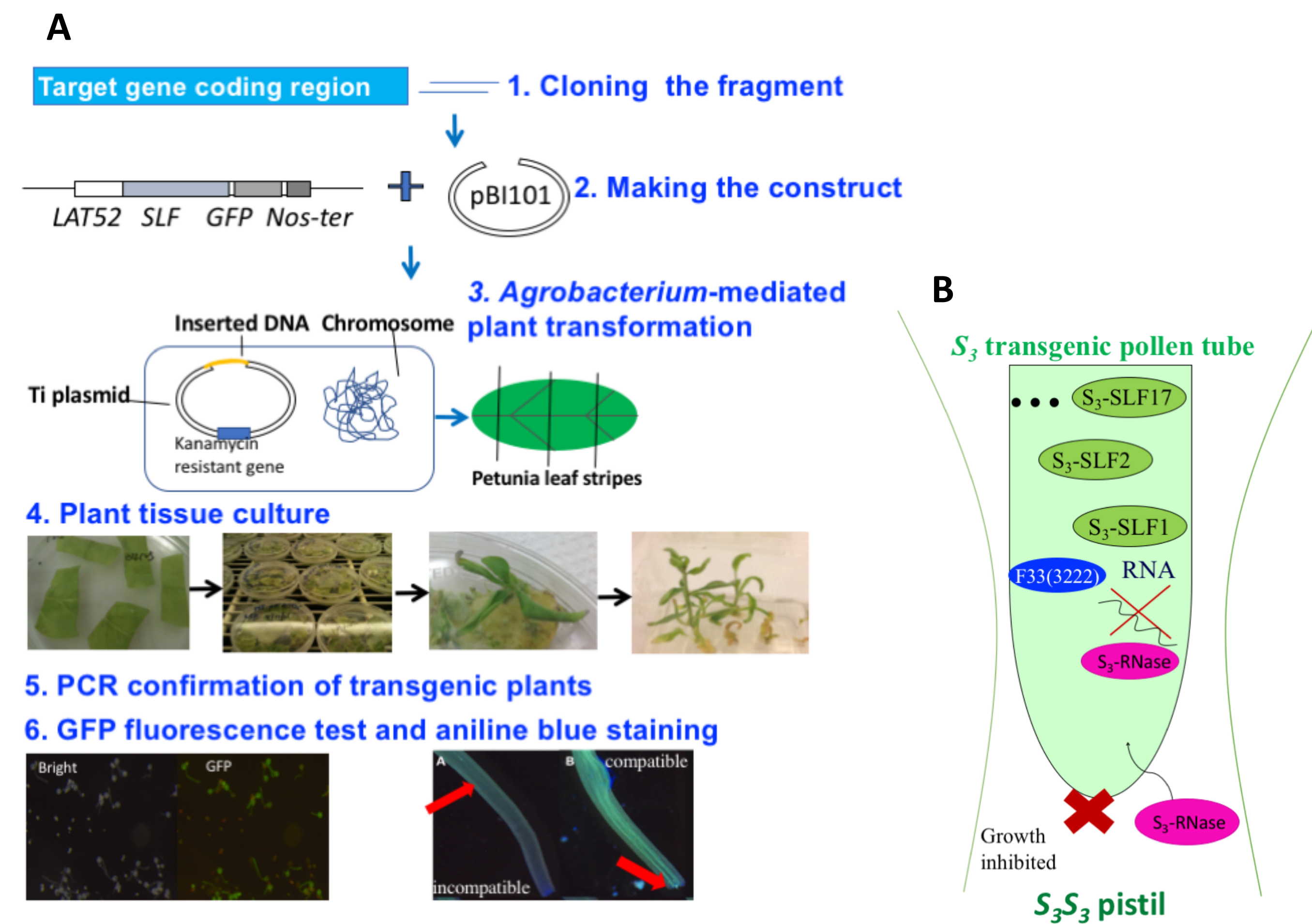


Figure 1. A. Standard procedure for generating transgenic plants expressing an *SLF* gene. B. *in vivo* functional assay

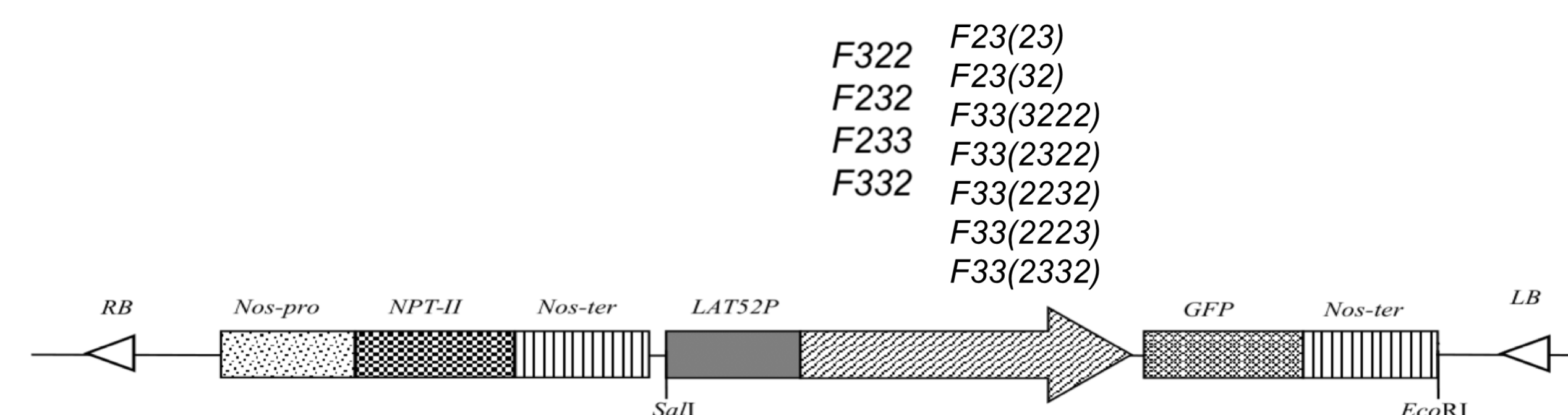


Figure 2. Transgene constructs for expressing 11 chimeric proteins of *S*₂-*SLF*1 and *S*₃-*SLF*1

Methods

To determine the biochemical basis for differential interactions of *S*₂-*SLF*1 and *S*₃-*SLF*1 with *S*₃-RNase, we first divided *SLF*1 into 3 functional domains (FD1, FD2 and FD3); generated 4 chimeric genes (*F*322, *F*232, *F*233, and *F*332) (Fig. 2); and used a transgenic assay (Fig. 1) to determine whether each encoded chimeric protein of *S*₂-*SLF*1 and *S*₃-*SLF*1 interacts with *S*₃-RNase. Based on the results (Fig. 3A), we further divided FD3 into 2 subdomains and then into 4 mini-domains (A, B, C, and D); generated 7 chimeric genes (Fig. 2); and similarly examined the ability of the resulting 7 chimeric proteins to interact with *S*₃-RNase (Fig. 3A,B). Protein structures were modeled using the I-TASSER server, and protein-protein docking analysis was performed by ClusPro (Fig. 4). FD3s of 4 *SLF*1s were aligned using MEGA 6 and ClustalW to identify amino acids conserved among the three that interact with *S*₃-RNase (Fig. 5).

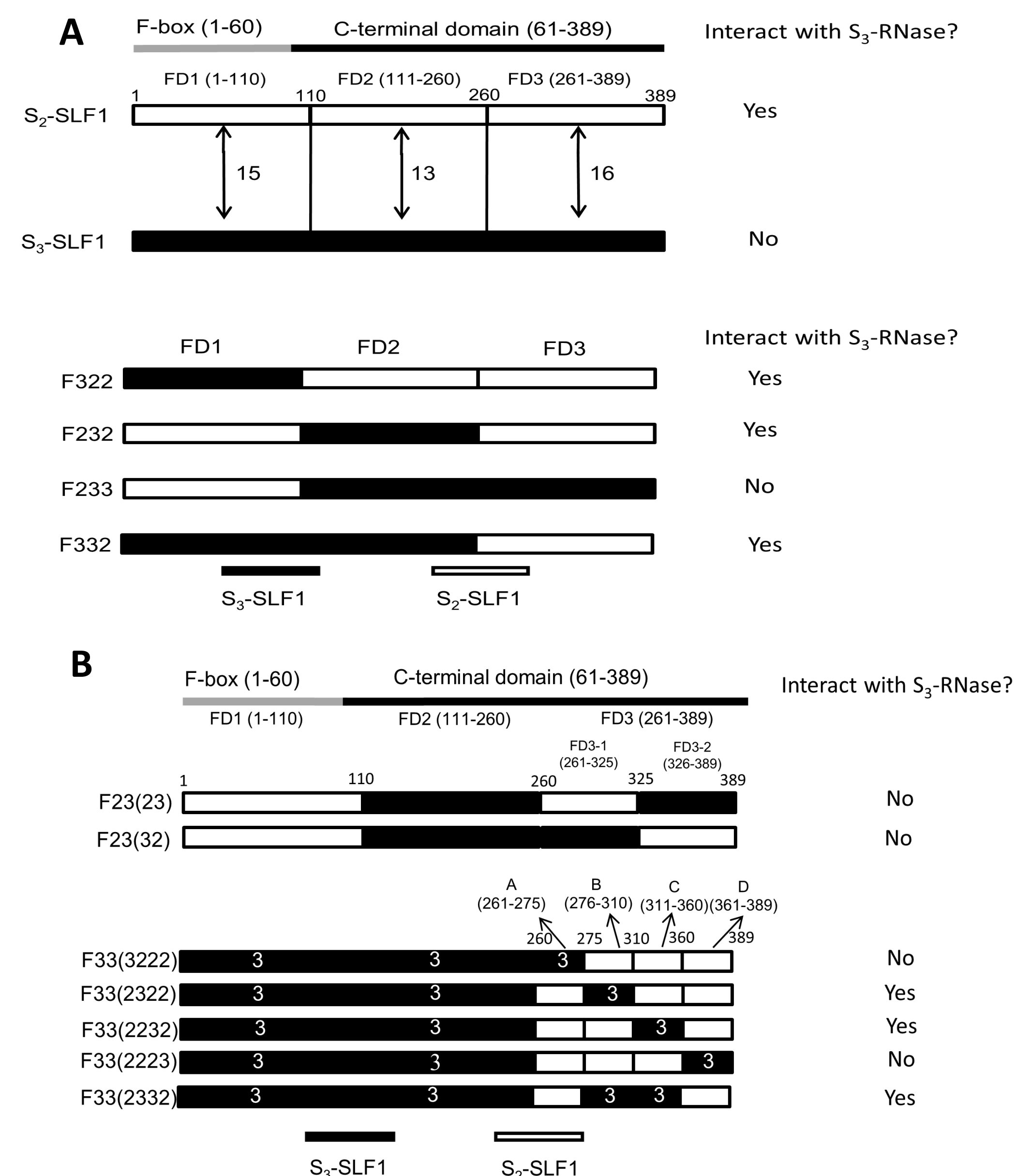


Figure 3. 11 chimeric proteins of *S*₂-*SLF*1 and *S*₃-*SLF*1, and their ability to interact with *S*₃-RNase.

Results

The results shown in Fig. 3 allowed us to first narrow the candidate amino acids for specific interaction of *S*₂-*SLF*1 with *S*₃-RNase to the 16 in FD3, and then to 4 in mini-domain A and 4 in mini-domain D. Molecular modeling of interactions between *S*₃-RNase and *S*₂-*SLF*1 revealed that 3 of these 8 are at the interaction surface (Fig. 4), and all 3 are conserved in *S*₁-*SLF*1 and *S*_{6a}-*SLF*1 that also interact with *S*₃-RNase (Fig. 5). Three of the chimeric proteins were used to determine whether FD3 alone contains the amino acids required for specific interaction of *S*₂-*SLF*1 with *S*₇-RNase and *S*₁₃-RNase. The results revealed that, unlike the case of *S*₂-*SLF*1's interaction with *S*₃-RNase, FD2 of *S*₂-*SLF*1 is required for interaction with *S*₇-RNase, and both FD1 and FD2 are required for interaction with *S*₁₃-RNase.

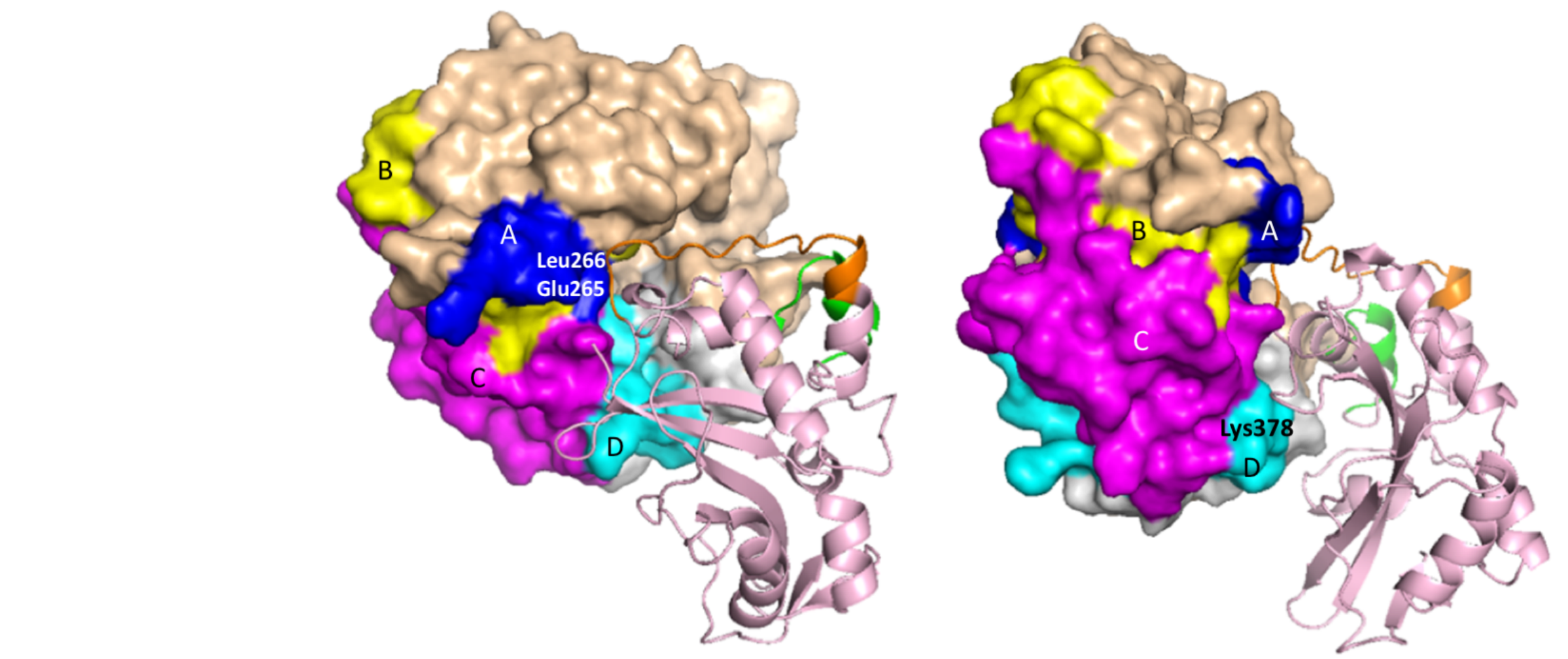


Figure 4. Computational modeling of *S*₂-*SLF*1 and molecular docking of *S*₃-RNase onto *S*₂-*SLF*1, as visualized in PyMOL.

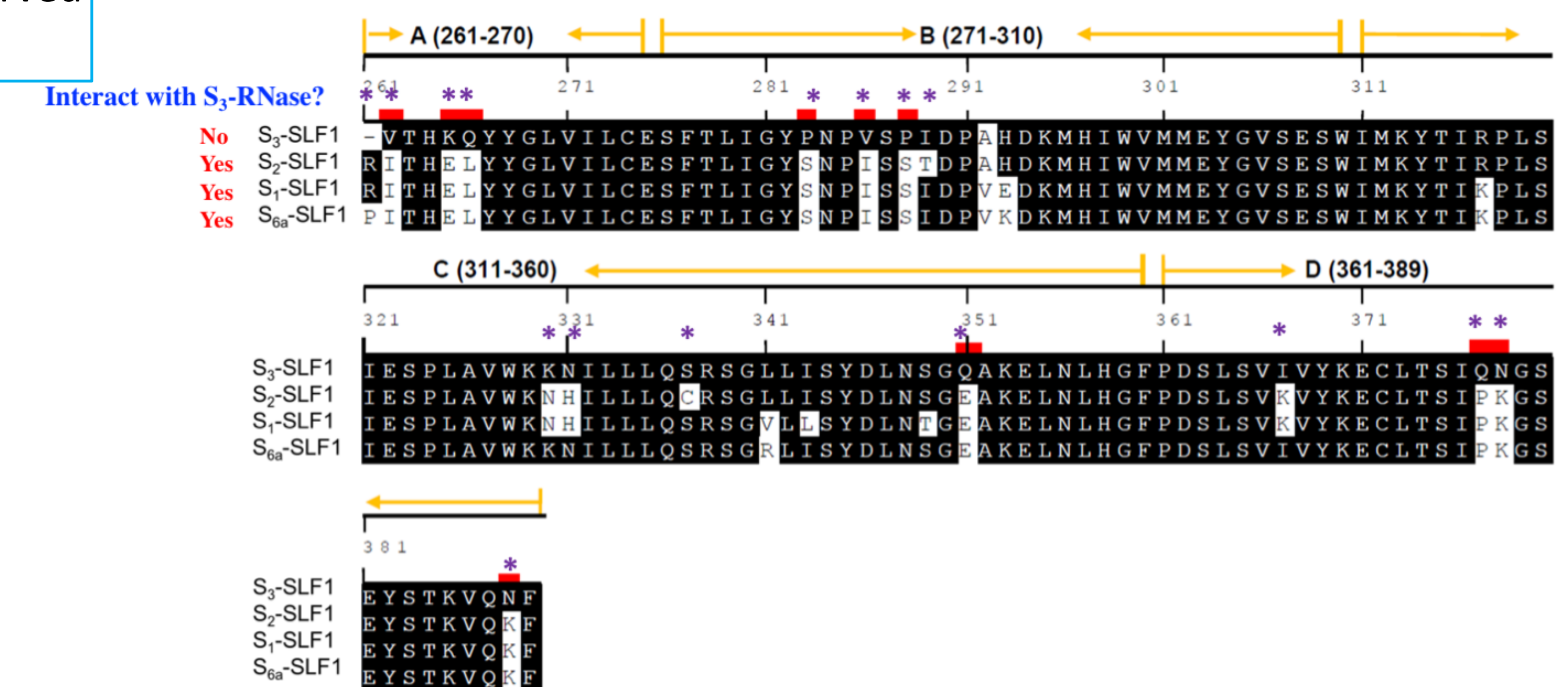


Figure 5. Alignment of deduced amino acid sequences in FD3s of three allelic variants of *SLF*1 that interact with *S*₃-RNase and of one allelic variant that does not.

Conclusion

Using domain-swapping and molecular modeling we have narrowed down the candidate amino acids for specific interaction of *S*₂-*SLF*1 with *S*₃-RNase from 44 that are different between *S*₂-*SLF*1 and *S*₃-*SLF*1 to 2 in mini-domain A and 1 in mini-domain D of FD3. In contrast, FD1, or both FD1 and FD2, contain(s) amino acids required for interactions with *S*₇-RNase or *S*₁₃-RNase, suggesting diversity and complexity of interactions between *SLF* proteins and *S*-RNases.

Acknowledgements

This work was supported by the National Science Foundation (IOS-1146182 and IOS-164557) to T.-h. K.

References

- Kubo K., Entani T, Takara A, Wang N, Fields AM, Hua Z, et al. (2010) Collaborative non-self recognition system in *S*-RNase-based self-incompatibility. *Science* 330: 796-799
- Williams JS, Natale CA, Wang N, Li S, Brubaker TR, Sun P, et al. (2014) Four previously identified *Petunia inflata* *S*-locus *F*-box genes are involved in pollen specificity in self-incompatibility. *Mol. Plant* 7: 567-569
- Wu L, Williams JS, Wang N, Khatri WA, San Roman D, Kao T-h (2018) Use of domain-swapping to identify candidate amino acids involved in differential interactions between two allelic variants of Type-1 *S*-locus *F*-box protein and *S*₃-RNase in *Petunia inflata*. *Plant Cell Physiol*. 59: 234-247